

Design, synthesis, structural characterization and cell cytotoxicity of a new derivative poly(maleic anhydride-co-vinyl acetate)/miltefosine polymer/drug conjugate

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In recent years, polymeric systems are selected as biomaterials because of their desired biocompatible properties and easy design/preparation of a number of different structures with lower toxicity and good solubility. Nontoxic polymeric drug carrier, maleic anhydride-co-vinyl acetate copolymer (MAVA), was prepared *via* free-radical chain polymerization at 80±0.1 °C. MEK (methyl ethyl ketone) and BPO (benzoyl peroxide) were used as the organic medium and radical initiator, respectively. Copolymer was conjugated with a broad-spectrum antimicrobial agent, miltefosine (MF, an oral drug in the treatment of leishmaniasis), Impavido[®] and Miltex[®], 1:1 molar ratio of copolymer:drug for 48 h at 60 °C in aqueous medium in presence of N-(3-dimethylaminopropyl)-N'-ethylcarbodiimide hydrochloride (EDAC). Fourier transform infrared (FTIR) and nuclear magnetic resonance (¹H- and ³¹P-NMR) were used to characterize the structure of the copolymer and MAVA/MF conjugate. Molecular weights were measured *via* size-exclusion chromatography (SEC). Results, obtained from the spectroscopic and SEC analysis, verified that conjugation was successfully carried out with good water-solubility. WST-1 cytotoxicity tests, 24 h by quantitative analysis, were carried out for copolymer, miltefosine, and MAVA/MF. The cytotoxicity values, by comparing with control group, were found statistically significantly different (P<0.05). MAVA/MF copolymer/drug couple was successfully designed with lower cytotoxicity than the free drug (MF).

Keywords: maleic anhydride-co-vinyl acetate copolymer conjugation, miltefosine, cytotoxic activity, FTIR, ¹H-NMR, ³¹P-NMR, SEC

INTRODUCTION

Anhydride containing polyanhydride copolymers have been commonly applied, specifically as solubilizing agents, nanoparticulate forms, surface modifiers, drug carriers, and also for other purposes such as diagnostic imaging agents and implants [1]. Generally, water-soluble conjugates could be well and easy designed as drug carriers for drug delivery. N-(2-hydroxypropyl)-methacrylamide (HPMA) copolymer-based drug delivery systems (DDSs) are recently used for clinical applications as anticancer agents. Biologically inert poly(ethylene glycol) (PEG), approved by Food and Drug Administration (FDA) for pharmaceutical trials that is preferred in many pharmaceutical, cosmetic, and industrial applications because of its water solubility, biocompatibility, nontoxicity, and non-immunogenicity characteristics [2]. Maleic anhydride (MA) copolymers mainly consist of MA and vinyl-based monomers. Modification/conjugation of the anhydride copolymers is accompanied by a chemical process in which the active anhydride unit can be opened after nucleophilic attack by the ring opening reaction

with amino (R-NH₂) or hydroxyl functional groups (R-OH) of nucleophilic reagents [3,4-6] to obtain amide/ester and carboxylic acid structures. We often encounter the amide/carboxyl structure because of its more favorable properties in our studies. Another main goal for this conjugation reaction was getting a copolymer/drug couple with better water solubility and lower cytotoxicity properties than the free drug for improvement of its main biological activity.

For more than 50 years, several techniques have been applied in pharmaceutical industry to combine biologically active agents with polymeric materials including PEG, poly(N-vinyl pyrrolidone), and cellulose derivatives [7]. Ringsdorf first proposed a model (schematized in Figure 1) known by its name in 1975 for design of polymer base prodrugs [8-10]. Commercially available vinyl base poly(styrene-co-maleic acid/anhydride) (SMA) is a well-known copolymer [3,11]. SMANCS conjugate consist of a SMA copolymer and the antitumor protein NCS (neocarcinostatin) [10,12-16].

The conjugate has also been applied in Japan for hepatocellular carcinoma treatment with significant therapeutic results and quite few side effects [17].

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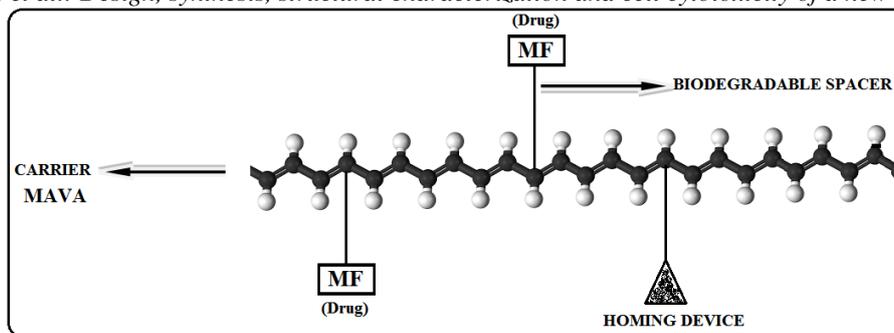


Figure 1. The Ringsdorf model.

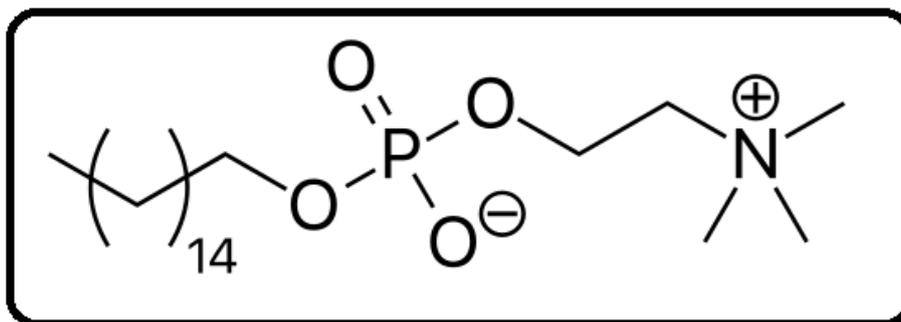
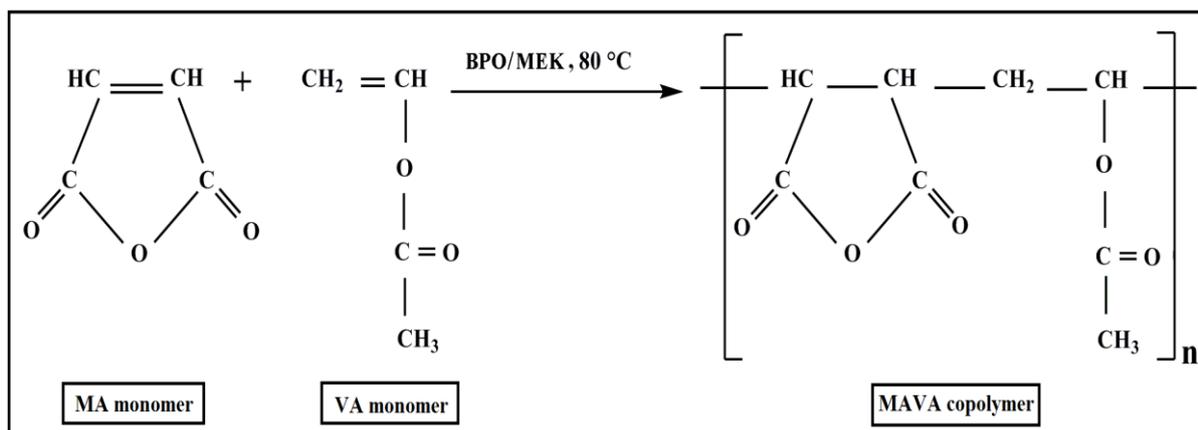
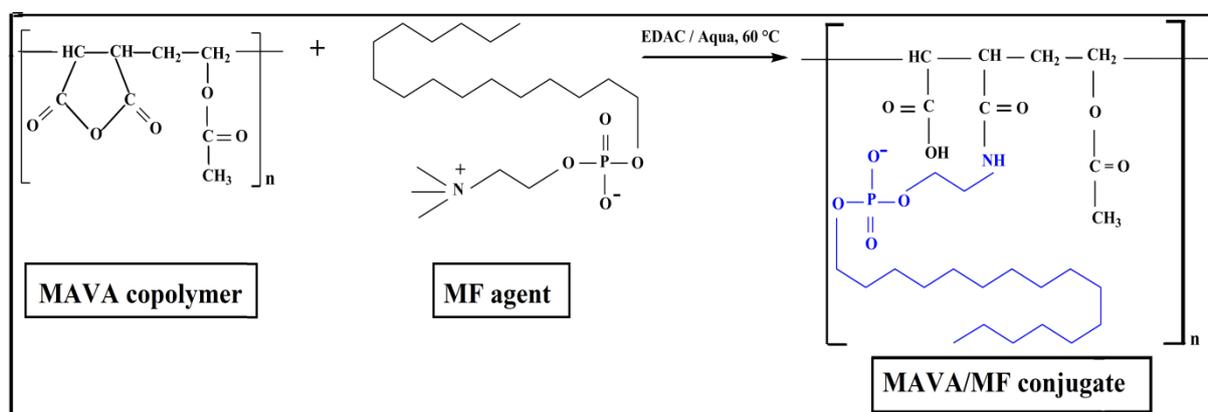


Figure 2. Chemical structure of miltefosine, MF, [2-(hexadecoxy-oxido-phosphoryl)oxyethyl-trimethyl-azanium].



Scheme 1. Polymerization reaction mechanism of the copolymer.



Scheme 2 Conjugation reaction of the copolymer with MF agent.

An antileishmanial drug, miltefosine [1-hexadecylphosphorylcholine (HePC)] is an

inhibitor of CTP: phosphocholine cytidyl transferase and has antimetastatic properties. It is

also more than 90% effective in eradicating visceral infections of *Leishmania* species. World Health Organization (WHO) reported that *Leishmaniases*, tropical diseases caused by the protozoan parasite *Leishmania*, are one of the six major diseases in developing countries [18]. Its systematic IUPAC name, chemical formula and molecular mass as follows:

2-(hexadecyloxy-oxide-phosphoryl)oxyethyl-tri-methyl-azanium, $C_{21}H_{46}NO_4P$, 407.568 g/mol. Miltefosine (MF) lateral chain consists of a phosphate (PO_4^{3-}) group, an intermediate ethyl chain ($-CH_2-CH_2-$), known with trade names Impavido[®], Miltex[®], a trimethylamine $N^+(CH_3)_3$ group and a terminated carbon side chain (Figure 2).

In this study, as a drug carrier maleic anhydride-vinyl acetate copolymer (MAVA) (Scheme 1) was chemically modified, through its reactive anhydride ring [19], by conjugation of the pharmaceutically active agent miltefosine, MF (Scheme 2). Conjugated product was named MAVA/MF. Cell cytotoxic effect of MAVA/MF was also studied with MAVA and MF cytotoxicity by using a healthy L929 mouse fibroblast cell line. Cytotoxicity experiments showed that MAVA/MF conjugate had no cytotoxicity when cell viability was taken into consideration even at 250 μ g/mL concentration.

EXPERIMENTAL

Materials

Synthesis, Conjugation and Purification: Chemicals were obtained from Merck (Schuchardt,

Germany) are as follows: MA (maleic anhydride), MEK (methyl ethyl ketone), EDAC [N-(3-dimethyl-aminopropyl)-N'-ethylcarbodiimide hydrochloride] and BPO (benzoyl peroxide). Ethyl alcohol (95%) was purchased from Carlo-Erba (Rodano, Italy). NaOH (sodium hydroxide), HCl (hydrochloric acid), VA (vinyl acetate), ethyl acetate and petroleum ether were ordered from Sigma-Aldrich (St. Louis, USA). Miltefosine was supplied by Orphanidis Pharma Research GmbH (Vienna, Austria).

Cultivation of L929 Mouse Fibroblast Cells and Cell Proliferation Assay: DMEM (Dulbecco's minimum Eagle medium) and fetal calf serum (10%) were obtained from Sigma (St. Louis, MO, USA), and WST-1 labeling reagent was purchased from Roche (Mannheim, Germany).

Synthesis and Purification of MAVA Copolymer

As described in our previous study [20], MAVA was synthesized traditionally by free radical chain polymerization reaction in the presence of MA and VA monomers. Table 1 summarizes the reaction conditions (briefly: 1:1 monomer molar ratio, MEK organic solvent, BPO free radical initiator, 24 h, and 80°C). Unreacted or homopolymer products were gently removed by incubating the white precipitated conjugate in ethyl acetate (24 h). Petroleum ether precipitation, under vacuum filtration, and vacuum incubator drying at 55 °C, 24 h were carried out for purification [21].

Table 1 Synthetic reaction conditions of copolymer and conjugated product.

SAMPLE	MOL PROPORTIONS	INITIATOR AND CATALYST	SOLVENTS	TIME (h)	TEMPERATURE (°C)
MAVA	MA:VA ~ (1:1)	BPO	MEK	24	80
MAVA/MF	MAVA:MF ~ (1:1)	EDAC	Water (ultra pure)	48	60

BPO: benzoyl peroxide, EDAC: N-(3-dimethyl-aminopropyl)-N'-ethylcarbodiimide hydrochloride.

Synthesis and Purification of MAVA/MF Conjugate

MAVA/MF conjugate was designed by the modification of MAVA copolymer with miltefosine (MF, mol. mass: 407.568 g/mol), at a 1:1 [6,22] molar ratio for 48 h at 60 °C in ultra-pure water, using EDAC [N-(3-dimethyl-aminopropyl)-N'-

ethylcarbodiimide hydrochloride] as the catalyst (activating role of carboxylic acid) (Table 1). Briefly, MAVA dried powder was hydrolyzed in alkaline water (pH 14), prepared in ultra-pure water using 1 M NaOH, and was then incubated by shaking continuously for 17 h at 50 °C in a dark

room for interacting all of the anhydride repeated units with concentrated NaOH [11]. Copolymer was expected to dissolve/hydrolyze in alkaline water medium and turn to a homogeneous transparent mixture which was immediately cooled again to room temperature, then the pH was reduced to 5 by drop-wise adding of 1 M HCl. Modified sample was prepared using the mixed solvent (5% ethyl alcohol/injectable water, stored at -20 °C until the adding step) [18] by drop-wise addition of ~10 mg/mL solution of MF. Initiation reaction was started by adding EDAC (20 mg/mL solution) over 15 min, pH was kept at 5 by adding 0.01 M HCl [11]. The reaction medium was then allowed to proceed for 24 h at 50 °C in a continuously shaking condition to get a viscous blurry white solution in a dark room, and allowed further for 48 h at 60 °C. The products were washed several times with excess of cold ethanol and incubated/cooled for 1h at -20 °C. White precipitate was then collected (centrifugation, 10 min, 3,000 rpm) in powder form after the liquid phase was removed and purified in a vacuum incubator (24 h, 55 °C) [23].

Structural Characterization

The infrared spectrum of the samples (copolymer and MAVA/MF conjugate) were recorded as KBr pellets on a FTIR spectrophotometer (Umicam MATTSON 1000, USA) at 400-4000 cm^{-1} with increments adjusted to 4 cm^{-1} . The nuclear magnetic resonance analyses, (^1H -NMR and ^{31}P -NMR) were performed on a Bruker Avance III instrument (Karlsruhe, Germany) at 400 MHz using 6 mg of the samples, dissolved in 0.8 mL of heavy water (D_2O).

Size-Exclusion Chromatography (SEC) of Copolymer/Drug Conjugate

As detailed in our previous study [23], average molecular weight distribution of the copolymer and modified product was measured by a Viscotek size exclusion chromatography (SEC) system (Houston, USA) with 660 nm refractive index and 670 nm right-angle light scattering (RALS) apparatus. Polystyrene standard was used for quadruple detector array calibration. Powdered samples were prepared in tetrahydrofuran (THF) and eluted (1.0 mL/min) at room temperature. OmniSEC 4.1 software program (Viscotek Cooperation, Houston USA) was used for obtained data analysis [24,25].

Cytotoxicity of MAVA Copolymer and MAVA/MF Conjugate

Cultivation of L929 mouse fibroblast cell lines: Detailed information about this method can be

obtained from our previous work [35]: L929 (ATCC cell line, NCTC clone 929) mouse connective tissue fibroblast cell line, was cultured in DMEM, in the presence of fetal calf serum (10%) and L-glutamine (2 mM/ml) without adding antibiotics to the cell culture medium. Cultivation was performed for the cultures in the humidified incubator at 37 °C and 5% CO_2 , until complete cell monolayer integration (~7 days).

Cell proliferation assay: As we clearly explained in our previous study [35], the cell proliferation assay mainly measures the number of living cells by dividing the tetrazolium salts to formazan by cellular enzymes in presence of the WST-1 labeling reagent. This method is based on a simple and well known kit protocol [35]. Briefly, after the user's kit protocol, quantification was performed by a scanning multi-well spectrophotometer (wavelength 450 nm) according to the absorbance measurement of the dye solution. Seeded cells in 96-well microtiter plates were adjusted to 1×10^5 cells/mL in a main volume of 100 μL per well. For the cytotoxic analysis of the MAVA; MF, and MAVA/MF 500, 250, 125, 62.5, 31.2, and 15.6 $\mu\text{g}/\text{mL}$ concentrations were applied. Incubation was performed for 24 h at 37 °C in a humidified atmosphere with 5% CO_2 . After the incubation period, WST-1 labeling reagent was gently supplemented to the medium as 1:1 volume ratio (10 μL) in each well. The absorbance values, using the control group comparison, were recorded at 450 nm using a Thermo Scientific microplate photometer (Multiskan FC, USA). Ten μL of WST-1 labeling reagent/one hundred μL of medium was also added to one well (selected blank position) on ELISA reader for background control purpose. After the start of the tetrazolium reaction (2 h) absorbance was recorded and experiments were carried out in six replicates for reliable results. Percentage viability was calculated according to user's manual and following Eq. (1) the optical density (OD) of the test products was compared with that of the negative control.

$$\text{Cell Viability (\%)} = \left[\frac{\text{OD}_{450} (\text{Sample})}{\text{OD}_{450} (\text{Negative control})} \right] \times 100 \quad (1)$$

Statistical analysis

Optical density (OD) results, with six replicates, were presented as mean \pm SD. Statistical ANOVA (one-way analysis of variance) was performed with 95% confidence levels for multiple comparisons. Furthermore, Student's *t*-test was also used for two group comparisons.

RESULTS AND DISCUSSION

FTIR Analysis

Unmodified copolymer (Figure 3a) had the main anhydride containing polymer structure [26-31]: symmetric and asymmetric carbonyl (C=O) stretches of MA at 1855.48 cm^{-1} , and 1780.59 cm^{-1} , respectively [26, 27]. Frequency vibrations of C-O-C bond on anhydride ring were found at 1025.21 cm^{-1} and 934.96 cm^{-1} [28]. Another carbonyl (C=O) symmetric and asymmetric stretches of VA was at 1715.67 cm^{-1} [29], CH_3 and CH_2 belong to VA stretching frequencies at 1373.80 cm^{-1} and 1430.38 cm^{-1} , respectively [30,31], characteristic acetyl (-COCH₃) group stretching frequencies of VA were at 1095.30 cm^{-1} [26], C-C vibration at 603.84 cm^{-1} , C-O-C stretching at 1214.91 cm^{-1} , and C-H group stretching vibration at 2943.19 cm^{-1} [32, 33].

Characteristic copolymer repeating units disappeared almost completely in MAVA/MF as a sign of the success of the ring-opening reaction (Figure 3b). It also means that a high percentage of the repeated anhydride rings were modified after copolymer hydrolysis and nucleophilic amino (-NH₂) group attack of MF [34]. Modification/conjugation of anhydride ring generally accompanied by functional group formation, for example, results in carboxylic group with amide or ester structure [34]. In this study conjugation reaction was verified with formation of a carboxylic acid and amide functionality. Characteristic -CH stretching vibration of VA along with the copolymer backbone stretching vibrations at 1403 cm^{-1} [35], CO-O-C ester groups or C-C of VA located at 1226.97 cm^{-1} , and a CH_2 deformation at 1585 cm^{-1} [36]. Peak observed at 1755 cm^{-1} could be assigned to -COOH carboxylic acid units [32]. 1394.31 cm^{-1} could be associated with MF because the same peak was also observed on pure MF spectrum. Peak 1066.28 cm^{-1} belongs to PO_4^{3-} antisymmetric stretching frequency [37-43], 1172.67 cm^{-1} and 1052.22 cm^{-1} were also assigned to PO_4^{3-} antisymmetric stretching movement [44]. According to literature, Breitinger and co-workers emphasized that the deceptively simple strong vibrations centered at 1059 cm^{-1} including minimum four components of inorganic phosphate

(PO_4), were produced by removal of the threefold degeneracy of PO_4 and activation of the general mode of phosphate (PO_4) [44]. Furthermore, 629.77 cm^{-1} was detected in the spectrum, because a series of infrared bands such as 620 cm^{-1} , 643 cm^{-1} and 673 cm^{-1} were assigned to out of vibrations of plane bending vibrational mode of the phosphate anion (PO_4^{3-}) and peak 892 cm^{-1} was associated with water vibrational modes [44]. Because the vibrational frequencies around 1577 cm^{-1} could be an overtone/combination band, the observed peak 1585.82 cm^{-1} was also considered as overtone and combination [44].

In addition to multiple phosphate vibrational modes detection, observed peaks were assigned to PO_4^{3-} , OH, and H_2O stretching and bending modes. Conjugate, MAVA/MF, consists of main polymeric backbone linked by PO_4 groups of miltefosine through an amide bond. As a result, at the molecular level there are phosphate units which are not equivalent in the conjugate structure. Peak located at 3662.75 cm^{-1} could be attributed to both primary amide and also carboxylic acid groups related to the most evident vibrational peak for conjugation reaction.

As reported previously by Das and co-workers, Figure 3c presents the IR spectra of the pure miltefosine. Peak assignments for the characteristics of MF: at 1471 cm^{-1} (bending vibrations for -CH₂) and 1242 cm^{-1} (asymmetric stretching vibrations for -PO₂) and overlap of 1078 cm^{-1} (symmetric stretching vibrations for -PO₂) and 1064 cm^{-1} (stretching vibrations for P-O-C) were detected in the solid state, crystalline form, of the drug MF [45].

NMR Analysis

¹H-NMR Analysis: ¹H-NMR spectrum of the copolymer (Figure 4a) showed the characteristic peaks as follows: chemical shift belongs to protons of anhydride rings at 3–4 ppm, -CH₂ protons on VA approximately at ~2.3–2.7 ppm, -CH fragment (bound directly to oxygen) produced a multiplet peak at around ~4.9–5.3 ppm [46, 47], and all of the -CH₃ protons, adjacent to C=O (carbonyl group), located at ~1.7–2.0 ppm [48-50].

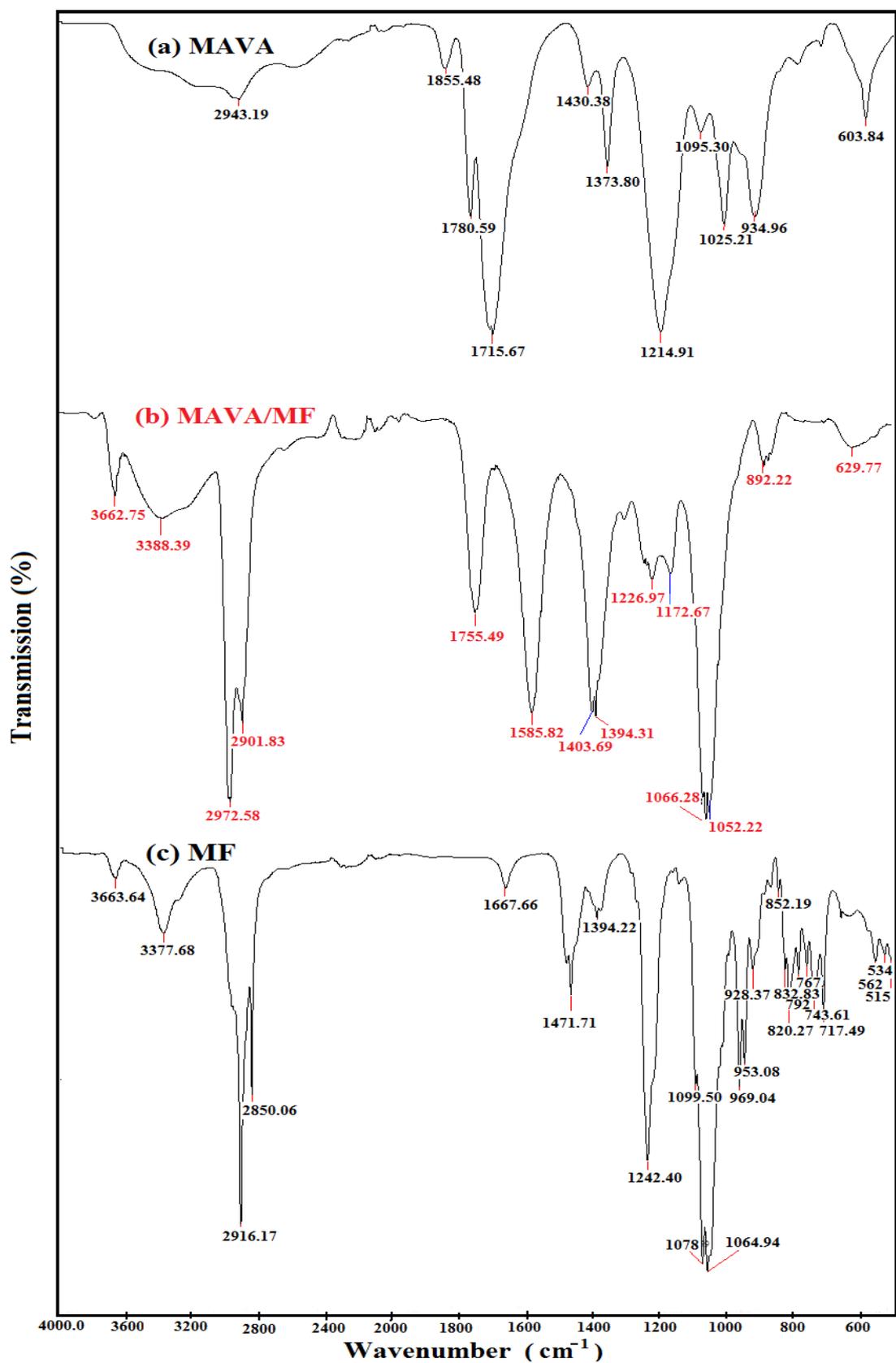
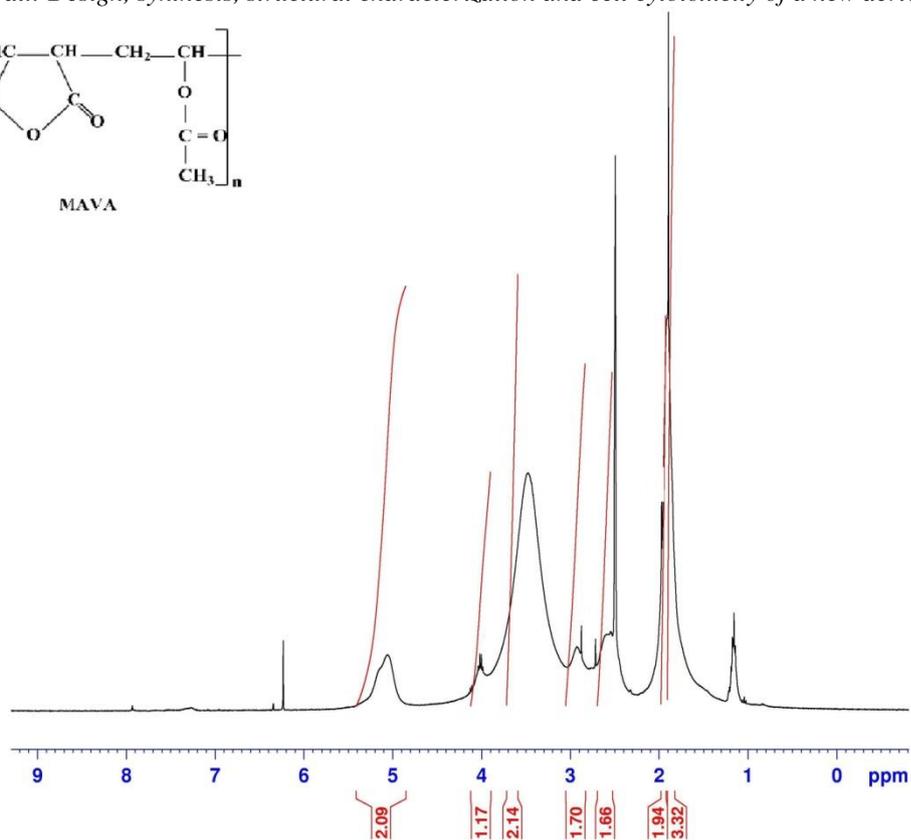
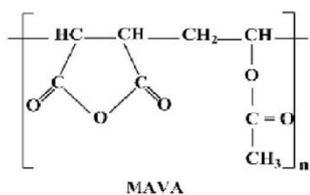
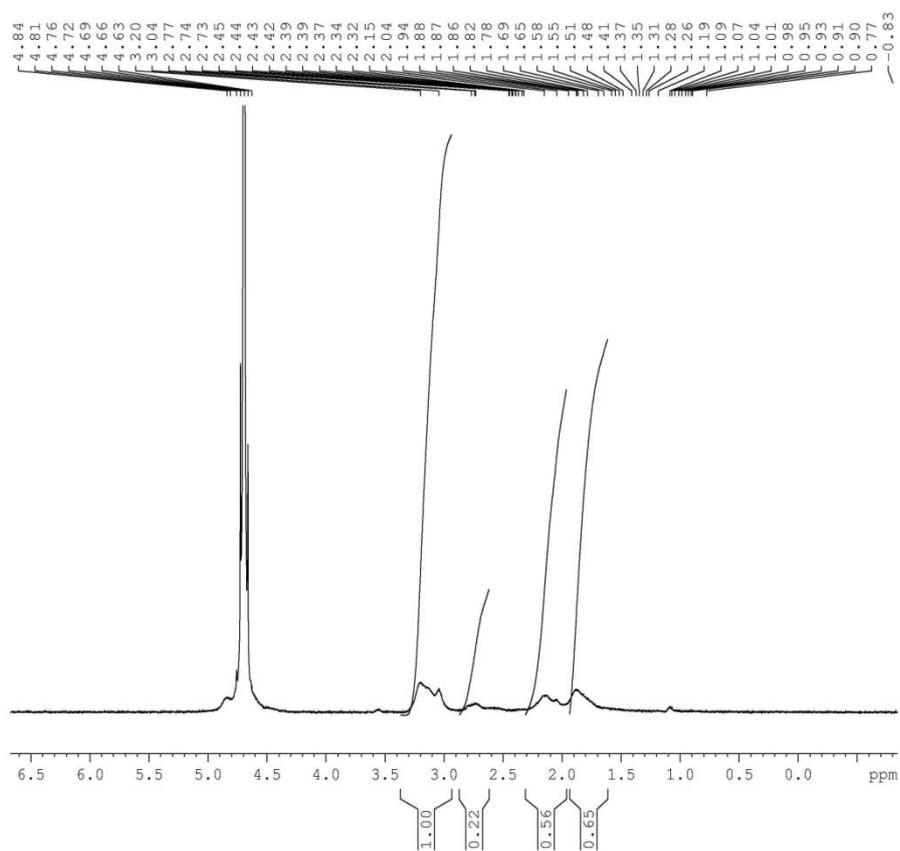


Figure 3. FTIR spectra of the copolymer, conjugation product, and MF agent.
 (a) MAVA, (b) MAVA/MF, and (c) MF



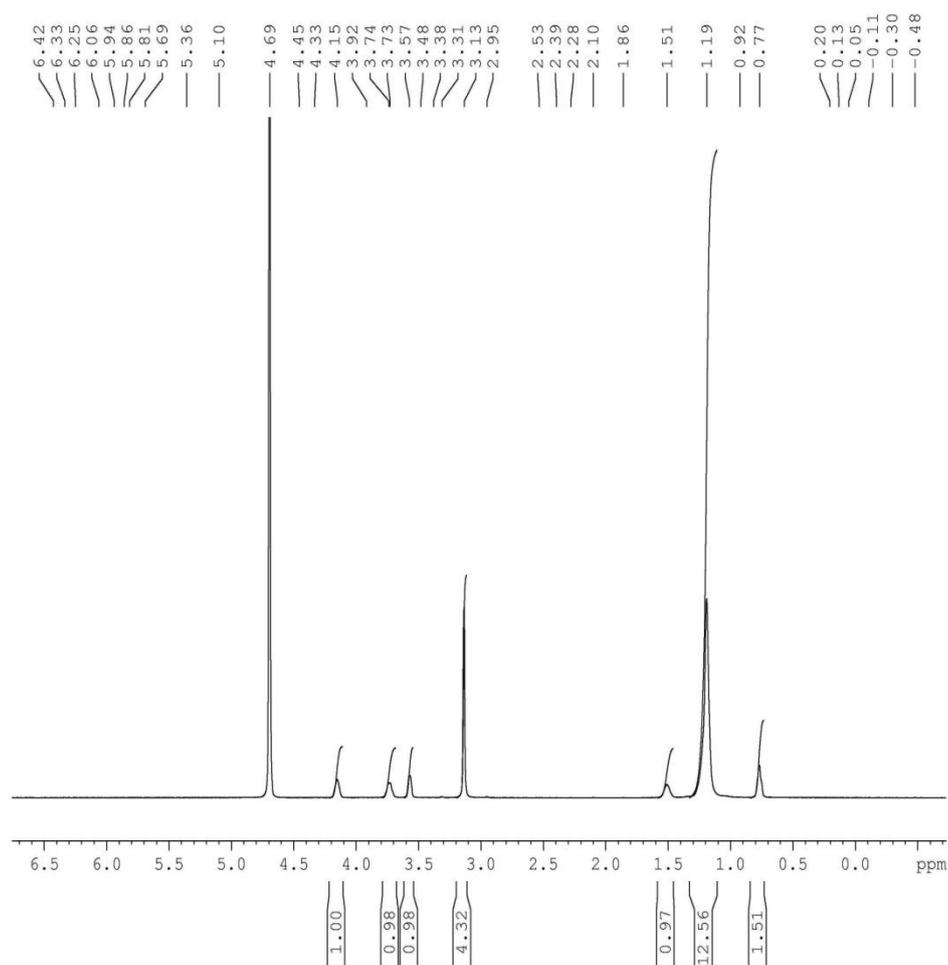
(a)

MAVA / MF



(b)

MF



(c)

Figure 4. $^1\text{H-NMR}$ spectra of the copolymer, conjugation product, and MF agent. (a) MAVA, (b) MAVA/MF, and (c) MF.

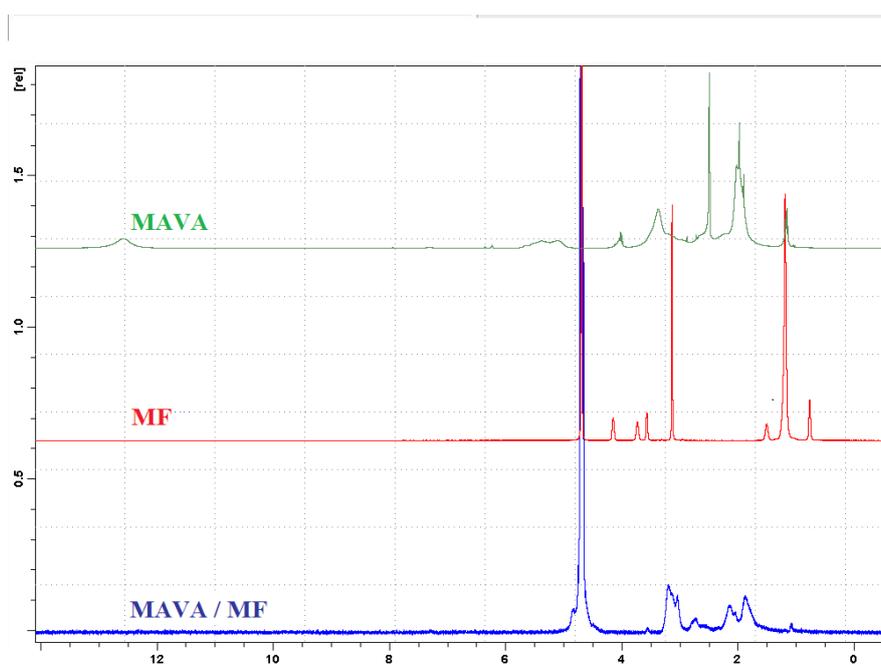
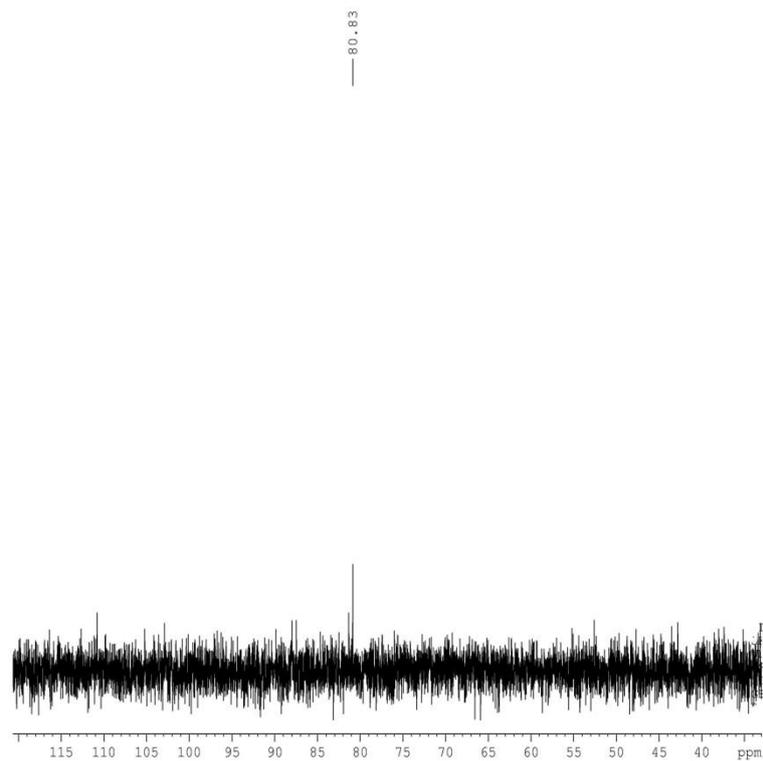


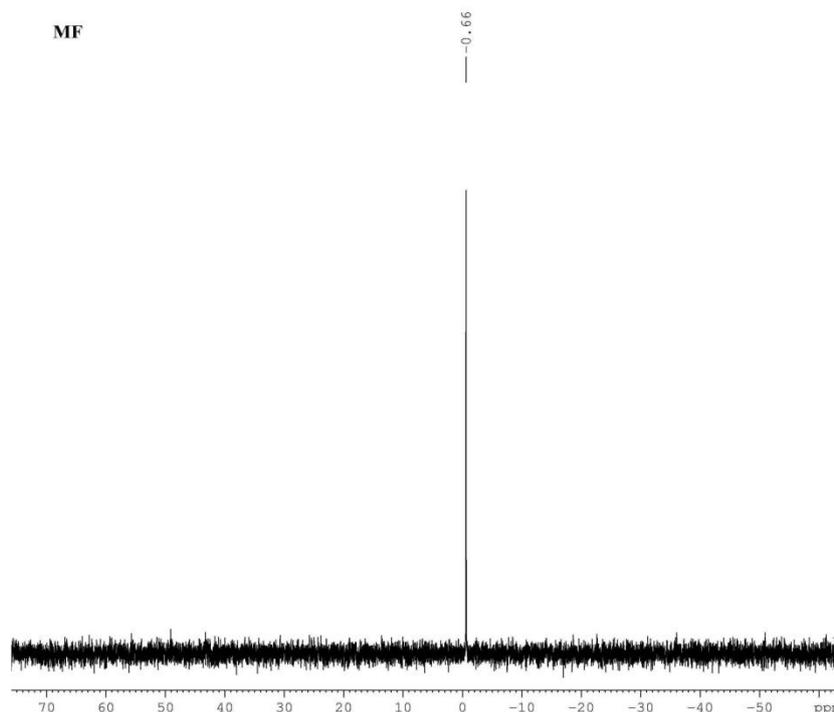
Figure 5. Overlapped $^1\text{H-NMR}$ spectra of the three components MAVA, MF and MAVA/MF.

MAVA/MF



(a)

MF



(b)

Figure 6. ^{31}P -NMR spectra of the conjugation product and MF agent.(a) MAVA/MF and (b) MF.

MAVA/MF conjugate (Figure 4b) peaks are as follows: aliphatic $-\text{CH}_3$ peaks of both copolymer backbone and end group of MF appeared at 2.77 ppm. It is taken into account that possible intramolecular hydrogen bonds with direct $-\text{NH}$ and $-\text{OH}$ fragments could not produce completely integrated peaks. Because the $-\text{NH}$ peaks of amide

linkage, evidence for the success of the chemical conjugation reaction, were masked/overlapped by the organic solvent peak at around 5.0–8.5 ppm, therefore, they were not detected. However, peaks observed at 4.84 ppm were assigned to amide linkage considering amide were overlapped by the solvent in the 4.63–4.84 ppm range with a slight

difference in chemical shift. D₂O solvent peaks were observed at 4.76 ppm [51] and 4.81 ppm (TSP). -CH₂ protons intensity on VA and MF appeared at 1.31–1.37 ppm. Peak range 2–3 ppm was attributed to the RNH-CH₂- (C is attached directly to N) as a marker of conjugation reaction, because the copolymer does not include any N atom. Due to the most important evidence of chemical modification of the copolymer with MF, 1.19, 1.26 and 1.28 ppm could be assigned to characteristic P-O-CH₂- linkage resulting from MF (Figure 4c) on the conjugate [52]. As can be clearly seen from the overlapped spectrum (Figure 5), both copolymer and MF chemical shifts were overlapped at around 1–5 ppm range. The most evident peak in the ¹H-NMR for the conjugation and also ring opening reaction of anhydride ring, shows the presence of phosphorus chemical shifts in the newly synthesized conjugate. All results proved that amide linkages and phosphorus on MAVA/MF were confirmed by ¹H-NMR and FTIR spectra. However, this conjugation reaction should be supported by a more detailed analysis like ³¹P-NMR analysis.

³¹P-NMR Analysis: Phosphorus-31 nuclear magnetic resonance (³¹P-NMR) is theoretically the same as ¹H-NMR spectroscopy. The ³¹P nucleus is useful in NMR spectroscopy due to its relatively high gyromagnetic ratio. Moreover, ³¹P has a 100% natural isotopic abundance property. Similar to the ¹H nucleus, the ³¹P nucleus has a nuclear spin of ±1/2 which makes spectra relatively easy and fast to interpret. As the phosphorus nucleus is coupled to a hydrogen nucleus bound directly to it, that is, this coupling separated by a single bond, is expected to be very high. Figure 6a shows the MAVA/MF spectrum, only one peak observed again at 80.83 ppm for PO₄⁻³ with some phosphorus chemical shift. It can be said that conjugation

reaction is verified by ³¹P-NMR due to phosphorus presence on the conjugate. Figure 6b shows a poor MF spectrum, only one peak observed at 0.66 ppm for PO₄⁻³ that is an evidence of phosphorus presence [53]. Overlapped ³¹P-NMR spectrum is also provided for the free and conjugated MF to compare both spectra (Figure 7).

It should be stressed that ³¹P-NMR is an excellent technique for studying phosphorus containing compounds, such as small organic compounds, metal coordination complexes, high molecular weight molecules, etc. Size-exclusion chromatography (SEC) was used for approximate molecular weights (Mw) measurement of pure copolymer and conjugated copolymer. Results obtained for MAVA copolymer: 3.98×10⁵ Da and MAVA/MF conjugate: 1.39×10⁶ Da (Da: daltons) are promising. SEC results confirmed that the molecular weight of conjugated copolymer with MF, reached almost threefold percentage, by the conjugation of MF to the copolymer backbone.

Supposed Reaction Mechanism for the Copolymer and Conjugate

Basic FTIR, ¹H-NMR and ³¹P-NMR spectroscopic analysis and obtained highly water-soluble characteristic of the conjugated product revealed that copolymer synthesized by free-radical polymerization and conjugation mechanism was in agreement with the assumption that the MF molecule was bound to the copolymer backbone by a ring-opening reaction (Schemes 1 and 2, respectively).

Average Molecular Weight Distribution of Copolymer/Drug Conjugate

Molecular weights of the MAVA copolymer and MAVA/MF conjugate were ~184 and ~548 gmol⁻¹, respectively.

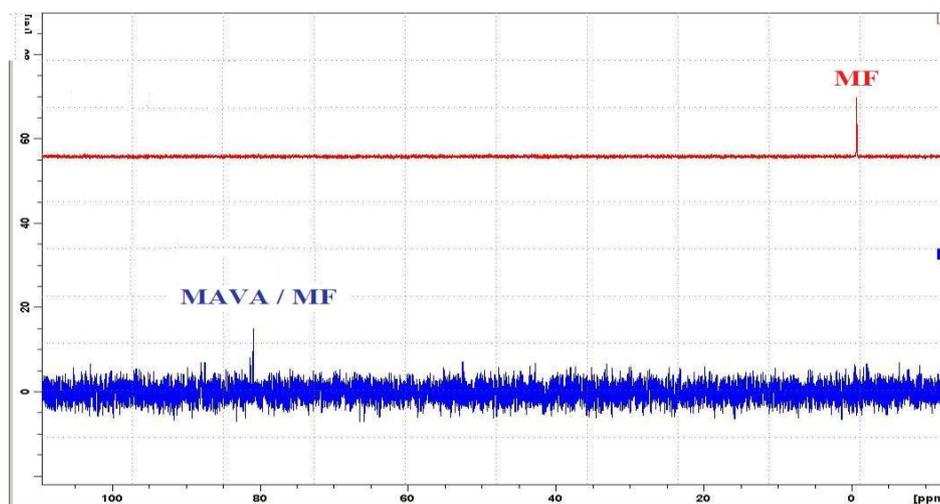


Figure 7. Overlapped ³¹P-NMR spectra of the two components MF and MAVAMF.

Cytotoxicity of agents: Quantitative analysis, according to WST-1 cytotoxicity test (24 h), were implemented for copolymer (MAVA), MF, and conjugate (MAVA/MF) (Figure 8).

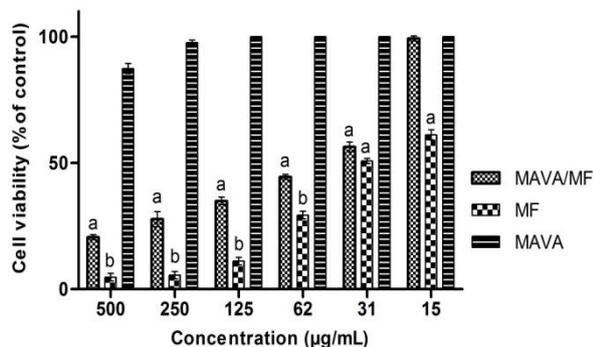


Figure 8. *In vitro* cytotoxic effect of the copolymer, MF, and MAVA/MF conjugate on L929 fibroblast cells lines. ^a P<0.05 vs control. ^b P<0.05 vs control, MAVA/MF 500, 250, 125 and 62.5 mg/mL.

The figure clearly shows that even at the highest concentration (500 µg/mL) of the copolymer, there was no adverse effect on living cells. The cytotoxicity results of MF and MAVA/MF 500, 250, 125, 62.5 mg/mL were compared with the control group for statistical evaluation, and were found significantly different (P<0.05) (Figure 8). It could be said for the cytotoxicity as follows: MF>MAVA>MAVA/MF (at 500, 250, 125, 62 and 31 mg/mL) and MF>MAVA=MAVA/MF (at 15 mg/mL). MAVA/MF copolymer/drug couple was successfully designed with less cytotoxicity than the free drug (MF).

CONCLUSIONS

To design a new copolymer/drug conjugate, nontoxic vinyl base copolymer (MAVA) was selected as the biocompatible drug carrier for MF. Chemical conjugation of the selected biologically active compound, MF (antileishmanial drug), was achieved by ring-opening reactions.

Potential toxicity measurement is generally used for the investigation of the viability effects of test compounds on the cells grown for a drug candidate. Synthetic polymers are considered to be promising materials for biomedical applications. Cell cytotoxicity assays of MAVA, MF, and MAVA/MF were carried out by quantitative WST-1 test (24 h). As a promising result it was observed that MAVA did not damage cell viability even at its highest concentration (500 µg/mL). The cytotoxicity results of MF and MAVA/MF 500, 250, 125, 62.5 mg/mL, by comparing with control

group, were found to be statistically significantly different (P<0.05).

Promising results should be emphasized, considering that MF is a highly toxic drug, that MAVA/MF had no cytotoxicity and is synthesized economically in a short time.

The initial goal of this experiment was to obtain a less/non-toxic drug having high water-soluble characteristics by comparing with free MF. On the other hand, preserving the original characteristics of miltefosine in the conjugate is a crucial step for rational copolymer/drug design.

In this study, in the direction of our main goal the conjugation reaction for copolymer/drug couple was successfully carried out with better water solubility and also less cytotoxicity than the free drug. We also hope that the biological activity of this conjugate (MAVA/MF) will be higher than that of the free drug (MF) with more comprehensive biological tests to be performed.

Acknowledgement: In this study, the polymeric samples, MAVA and MAVA/MF, were synthesized in the Research Center (CUTFAM), Cumhuriyet University, Faculty of Medicine (Sivas, Turkey). FTIR and NMR analysis were carried out at the Technology Research and Developing Centre (TAUM), Erciyes University (Kayseri, Turkey). Molecular weight measurements of the samples were performed by SEC (Size-Exclusion Chromatography) analysis at Yildiz Technical University (Istanbul, Turkey).

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